



## Original Research Article

# Isolation, Characterization and Estimation of Antimicrobial Activity of Novel Bacteriocin from *Lactobacillus plantarum*

Arunava Das\*, Santanu Sasidharan, Thejus Achuthan, M.E.Sindhuja

Department of Biotechnology, Bannari Amman Institute of Technology,  
Sathyamangalam-638401, Tamilnadu, India

\*Corresponding author

## ABSTRACT

### Keywords

Food borne pathogens;  
*Salmonella typhi*;  
*Bacillus subtilis*

Food borne pathogens are becoming a matter of great concern in various industries like dairy, egg and other food industries. Various gram positive and gram negative bacteria like *Salmonella typhi*, *Bacillus subtilis* are the main causative organisms. The present study is based on isolation, identification and screening of major food borne pathogens and novel bacteriocin producing strain of *Lactobacillus plantarum*. Investigation was conducted on 389 food samples and 12 different genus of bacteria were identified and isolated. From the 62 isolates of *Lactobacillus* sp. isolated, 22 isolates were *Lactobacillus plantarum* and of which 14% (3 isolates) were found capable of producing bacteriocin in large amounts. The isolates were grown for 96 hours and the bacteriocin was extracted by centrifugation. Ammonium sulphate precipitation at 70% level of saturation was performed and the precipitate was centrifuged. Well diffusion assay of the extracted protein was performed on different isolated food borne pathogens and the diameter of the zones of inhibition was recorded. The study reveals the extensive scope of bacteriocin in the field of food industry as preservatives and the need for further research in the field

## Introduction

The risk of contamination by pathogens mostly food borne microorganism is becoming a matter of great concern in the day to day world. This decreases the quality control of the food products and thereby decreasing consumer demand for the product in the market. The Food and Drug Administration (FDA) have prioritized the matter as top level and have listed the food borne pathogens in

Bacteriological Analytical Manual, FDA (FDA, 2012). The list include various microorganisms like *Escherichia coli*, *Salmonella* sp., *Shigella* sp., *Listeria monocytogenes*, *Staphylococcus aureus*, and *Bacillus subtilis*. The pathogens recorded previously of causing contamination in food industry are *Staphylococcus* and *Streptococcus* species. Bacteriocins are classified as antibacterial

peptides or proteins that are synthesised by bacteria as a microbial defence mechanism. At the molecular level, they are synthesized by small ribosomes and they can permeabilize through membrane and are cationic in nature (Klaenhammer<sup>a</sup>, 1993; Jack *et al.*, 1995; Thompson *et al.*, 1996). Previous studies have found the antimicrobial nature of the peptides. Despite of their different source, structure, mode of action and specificity, any molecule of protein that is secreted by the bacteria and has antimicrobial activity is considered to be a bacteriocin (Rammelsberg and Radler, 1990). These bacteria inhibits gram positive bacteria and food spoilage bacteria (Klaenhammer<sup>b</sup>, 1988, Caslaet *et al.*, 1996, Ennanet *et al.*, 1996, Contreras *et al.*, 1997, Messiet *et al.*, 2001) and gram negative bacteria ( Lewuset *et al.*, 1991, Stevens *et al.*, 1991, Messiet *et al.*, 2001).

Lactic acid bacteria (LAB) are characterised as gram positive bacteria and they are cocci or rod shaped. The genus is anaerobic but can tolerate and grow in the presence of air. These bacteria produce antagonist substances called bacteriocins which have high antimicrobial activity in low concentration (Klaenhammer<sup>a</sup>, 1993, Moronoet *et al.*, 2006), The antimicrobial activity of these compounds have increased the scope of research and interest in the isolation of *Lactobacillus sp.* producing bacteriocin and characterisation of these peptides (Derazet *et al.*, 2005).

## Materials and Methods

### Bacterial Strains Isolation and Culture Conditions

389 samples were collected from randomly selected from various retail

shops in Erode, Tiruppur, Namakkal and Coimbatore districts of Tamil Nadu, India and immediately to laboratory conditions for isolation. The various samples procured were meat, fish products, milk, dairy products, raw vegetables, bakery products, beverage and fermented rice products. The food samples were aseptically inoculated into freshly made and sterile Brain Heart Infusion broth (Hi-Media Laboratories, Mumbai) test tubes and is maintained aerobically at fermented material and the food samples from BHI broth were inoculated into culture specific medium De man Rogosa Sharpe (Hi-media Laboratories), Tryptone Soy Agar (Hi-media Laboratories), Sheep Blood Agar (Hi-media Laboratories), MacConkey Agar (Hi-media Laboratories), Xylose lysine deoxycholate Agar (Hi-media Laboratories) and incubate characters were selected randomly and repeated streaking in fresh agar culture was carried out each time until pure culture is obtained. The pure cultures were regrown in Nutrient Agar characterisation according to Bergey's manual of determinative bacteriology (Holt *et al.*, 1994).

### Screening of Isolates for Bacteriocin Production

The bacteriocin produced by 22 isolates of *Lactobacillus plantarum* was checked for activity by screening against maximum amount of food borne pathogens isolated. The isolates of anaerobically. The culture was centrifuged at 10000xg for 15 mins and the supernatant was collected. The crude bacteriocin was tested for activity by well diffusion method. The well dimensions was maintained at 7mm in diameter and 5mm deep in each culture and 35µl sample was added to each vessel. The diameter of the zones were

**Table.1.1** Morphological Characteristics of isolated bacteria (Suspected)

Bacteria Investigated	Motility Test	Gram Staining	Flagella Staining	Endospore Staining
<i>Aeromonassorbia</i>	Motile	Gram Negative	Single Polar Flagella	No Endospore
<i>Bacillus cereus</i>	Motile	Gram Positive, Rod	Peritrichous Flagella	Central
<i>Bacillus subtilis</i>	Motile	Gram Positive, Rod	Peritrichous Flagella	Subterminal
<i>Escherichia coli</i>	Motile	Gram Negative, Rod	Peritrichous Flagella	No Endospore
<i>Klebsiellaoxytoca</i>	Non-Motile	Gram Negative, Rod	No Flagella	No Endospore
<i>Klebsiella pneumonia</i>	Non-Motile	Gram Negative, Rod	No Flagella	No Endospore
<i>Listeria monocytogenes</i>	Motile	Gram Positive, Cocci	Peritrichous Flagella	No Endospore
<i>Salmonella enterica</i>	Motile	Gram Negative, Rod	Peritrichous Flagella	No Endospore
<i>Staphylococcus aureus</i>	Non-Motile	Gram Positive, Cocci	No Flagella	No Endospore
<i>Streptococcus agalactiae</i>	Non-Motile	Gram Positive, Cocci	No Flagella	No Endospore

**Table.1.2** Percentage of Isolated Tested Positive for Biochemical Tests

Bacteria Investigated	No. of Isolates	NR	SH	CT	CA	OX	UR
<i>Aeromonassorbia</i>	34	97	-	85	-	84	-
<i>Bacillus cereus</i>	22	92	89	100	85	-	59
<i>Bacillus subtilis</i>	31	*1	86	99	90	64	-
<i>Escherichia coli</i>	86	100	-	-	85	-	-
<i>Klebsiellaoxytoca</i>	12	95	-	99	80	-	89
<i>Klebsiella pneumonia</i>	21	95	-	98	84	-	85
<i>Listeria monocytogenes</i>	82	-	-	-	99	-	-
<i>Salmonella enterica</i>	8	95	-	-	88	-	-
<i>Staphylococcus aureus</i>	11	100	-	99	85	-	-
<i>Streptococcus agalactiae</i>	54	-	-	-	-	-	-

NR-Nitrate Reduction, SH-Starch Hydrolysis, CT-Citrate, CA-Catalase, OX-Oxidase, UR-Urease

**Table.2.1** Antimicrobial Activity of Crude and Partially Purified Bacteriocin

Isolated Strains	Zone of Inhibition (Crude)mm			Zone of Inhibition(Partially Purified)mm		
	LP-FP-6	LP-BR-3	LP-CD-12	LP-FP-6	LP-BR-3	LP-CD-12
<i>Aeromonassorbia</i>	6	6	5	7	9	7
<i>Bacillus cereus</i>	7	8	7	10	10	11
<i>Bacillus subtilis</i>	8	8	7	13	12	10
<i>Escherichia coli</i>	6	5	4	9	9	9
<i>Klebsiellaoxytoca</i>	0	0	0	0	0	0
<i>Klebsiella pneumonia</i>	2	3	2	5	8	5
<i>Listeria monocytogenes</i>	6	8	6	7	10	9
<i>Salmonella enterica</i>	2	5	2	5	8	4
<i>Staphylococcus aureus</i>	9	12	10	13	13	12
<i>Streptococcus agalactiae</i>	5	8	5	8	9	8

recorded for maximum number of indicator bacterial isolates to select the best strains.

#### Extraction of Bacteriocin:

250 ml of MRS broth was inoculated with the best strains of *Lactobacillus plantarum*

#### Antimicrobial Assay of purified bacteriocin

Antimicrobial property of the purified bacteriocin against various food borne pathogen isolates by well diffusion assay (Sharma *et al.*, 2011). 1 ml of the indicator bacteria was swabbed on a previously poured and sterilised nutrient agar plates using sterile cotton buds.

### Results and Discussion

#### Isolation and Identification of Bacteria

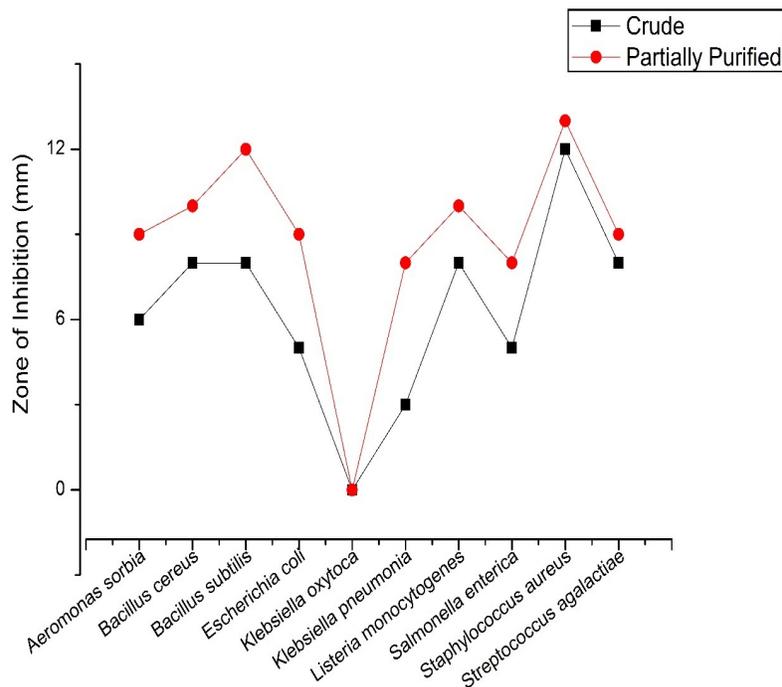
From a total of 389 samples were investigated and from which a total of 688

species were isolated. The suspected bacterial colonies were purified by repeated streaking in selective agar media plates until the pure cultures were obtained. The morphological and biochemical characteristics were recorded.

#### Antimicrobial Assay

Antimicrobial assay was performed with both crude and partially purified bacteriocin. The partially purified bacteriocin displayed a large zone of inhibition in microorganisms like *Staphylococcus aureus* and *Bacillus aureus*. The results were almost similar to the study done by Dhanpathiet *al.*, 2008. The bacteriocin produced by *lactobacillus plantarum* was found to be effective against both gram positive and gram negative bacteria. The comparison of crude and purified bacteriocin revealed an increase in activity. The study reveals a positive co-relationship with the study done by Bizaniet *al.*, 2002.

**Figure.2.2** Comparison of Zones of Inhibition of Crude and Partially Purified



The presence of various microorganism like *Aeromonas*, *Bacillus*, *Escherichia*, *Klebsiella*, *Listeria*, *Staphylococcus*, *Salmonella* and *Streptococcus* was confirmed but the present study in different food, fermented and dairy products. The strains of *Lactobacillus plantarum* which produces bacteriocins were isolated and the crude and partially purified bacteriocins were tested for antimicrobial activity successfully. The inhibition was found to be more active in gram positive bacteria and the prospects of the usage of these bacteriocins in the food industry as preservatives is large. Further studies like large scale production and structural analysis can be implored on and way could be paved for profound research

### Acknowledgement

The authors wish to acknowledge the financial grant with reference no.

20/AICTE/RIFD/RPS (POLICY-1)28/2012-13 under Research Promotion Scheme of All India Council of Technical Education (AICTE), New Delhi to execute this research work as a part of the funded project. Authors would also like to convey thanks to Bannari Amman Educational Trust for the kind support.

### References

- Bizani, D. and Brandelli, A. 2002 'Characterization of a Lisboa and bacteriocin produced by a newly isolated *Bacillus* sp. strain 8', A.J.Appl.Microbio., Vol93, pp.519-523
- Casla, D., Requene, T. and Gomez, R. 1996 'Antimicrobial activity of lactic acid bacteria isolated from goat's milk and artisanal cheeses: characteristics of a bacteriocin produced by

- Lactobacillus curvatus* IFPL 105', J.Appl.Bacteriol., Vol.81, pp.35-41.
- Contreras, B.G., De Vuyst, L., Devreese, B., Busanyova, K., Raymaeckers, J., Bosman, F., Sablon, E., and Vandamme, E.J., 1997 'Isolation, purification and amino acid activity of *Bacillus subtilis* extract on pathogenic organisms', Tamil Nadu J.Veterinary and Anim. Sci., Vol.4, No.4, pp.150-153
- Jack, R.W., Tagg, J.R. and Ray, B. 1995 'Bacteriocins of Gram Positive Bacteria' Microbiol Rev., Vol.59, pp.171-200
- Klaenhammer, T.R. 1993 'Genetics of bacteriocins produced by lactic acid bacteria', FEMS Microbiol. Rev. Vol.12, pp.39-86
- Klaenhammer, T.R. 1988 'Bacteriocins of lactic acid bacteria', Biochimie. Vol70, pp.337-349.
- Messi, P., Bondi, M., Sabia, C., Battini, R. and Manicardi, G. 2001 'Detection and preliminary characterisation of a bacteriocin plantaricin 35d produced by *Lactobacillus plantarum* strain', Int.J.FoodMicrobiol., Vol.64, pp.193-198
- Moreno, I., Lerayer, A.S.L., Baldini, V.L.S. and Leitao, M.F.de F. 2000 'Characterization of bacteriocins produced by *Lactococcus lactis* strains' Braz. J. Microbiol., Vol.31, pp.184-192
- Rammelseberg, J.M 1996 'Review: Antimicrobial spectrum, structure, properties and mode of action of nisin, a bacteriocin produced by *Lactococcus lactis*', Food Sci. Technol. Int., Vol.2, pp.61-68
- Sharma, N., Riti, K., Neha, G and Ranjana, K. 2011 'Purification of Bacteriocin by *Bacillus subtilis* R75', Food Technol. Biotechnol., Vol.49, No.2, pp.169-176.
- sequencing of lactobin A, one of two bacteriocins produced by *Lactobacillus amylovorus* LMG p-13139', Appl.Environ.Microbiol., Vol.63, pp.13-20.
- Dhanapathi., Prabhakar, T.G. and Prabakar, P.2008 'Antibacterial Thompson, J.K., Collins, M.A. and Mercer, W.D. 1996 'Characterization of a proteinaceous antimicrobial compound produced by *Lactobacillus helveticus* CNRZ 450', J.Appl. Bacteriol., Vol.80, pp.338-348